NucleoSpin® Plasmid – isolation of M13 DNA (Rev. 01)

This protocol is only a supplement to the kit's general user manual. Please refer to the kit manual for more detailed information regarding safety instructions, product-specific disclaimers, and especially preparations needed before starting the procedure. The latest version of the user manual is available at ***www.mn-net.com/usermanuals*** *or can be requested from our technical service (tech-bio@mn-net.com). Material safety data sheets (MSDS) can be downloaded from* ***www.mn-net.com/MSDS****.*

Additional reagent needed:

- Buffer NT2 (REF 740597)
- Glacial acetic acid

1 Cultivate and harvest bacterial cells

Grow a M13-infected *E. coli* culture and centrifuge bacterial cells at **4,000 x g** for **10 min** at **4 °C**.

2 Cell lysis

Transfer the supernatant to a new microcentrifuge tube. For each **1 mL supernatant** (containing suspended phage particles), add **10 µL glacial acetic acid**. Mix by inverting the tube 6–8 times. Incubate at room temperature for **2 min**.

Place a NucleoSpin® Plasmid Column into a Collection Tube (2 mL) and load sample. Centrifuge for **1 min** at **11,000 x *g*** and discard flow-through.

Maximal loading volume of a NucleoSpin® Plasmid Column is 700 µL. If larger volumes are to be processed, load samples in successive steps. Do not load the column more than 3 times.

Place the NucleoSpin® Plasmid Column back into the collection tube and add **700 µL** of **Buffer NT2**. Centrifuge for **1 min** at **11,000 x *g*** and discard flow-through.

Place the NucleoSpin® Plasmid Column back into the collection tube, add 700 µL of Buffer NT2, and incubate for 1 min at room temperature. Centrifuge for 1 min at 11,000 x ***g*** and discard flow-through.

3 Isolate M13 DNA

Continue with step 5 ('Wash silica membrane') of the NucleoSpin® Plasmid standard protocol.